

RELATIONSHIP BETWEEN PLANT LIPID BODIES AND FUNGAL ENDOPHYTES

Relación entre Cuerpos de Lípidos y Hongos Endofitos

Pedro Osuna Avila^{1‡}, Jerry R. Barrow², Mary E. Lucero², and Ronald E. Aaltonen²

SUMMARY

Lipid bodies are universal components of plant cells and provide a mobilized carbon source for essential biological processes. Oil plants harvested for food and fuel depend on these lipid bodies. Plants also host diverse populations of endophytic fungi, which easily escape microscopic detection. This study reviews data from previous surveys of endophyte distribution in native plants to specifically examine the physical association between endophytic fungi and plant lipid bodies. Plant tissues stained with trypan blue and sudan IV prior to differential interference contrast microscopy exhibited lipid bodies tightly associated with fungal hyphae and with trypan blue stained fungal networks. The abundance of endophyte-associated lipids in healthy plant tissues suggests endophyte involvement in carbon (oil) metabolism and transport. More research, particularly at the molecular level, is merited to assess the significance of this plant feature which is conserved in grasses and shrubs. Exploring the interfaces where plant cells and endophyte cells exchange organic carbon by using modern imaging, molecular and genomic analysis could transform current understanding of plant carbon metabolism.

Index words: arid zones, biofuel, endophyte, fungi, stress resistance.

RESUMEN

Los cuerpos de lípidos son componentes universales de las células vegetales que conllevan una fuente de carbono que es esencial para los procesos biológicos.

¹ Universidad Autónoma de Cd. Juárez. Henry Dunant 4016. 32310 Cd. Juárez, Chihuahua, México.

[‡] Autor responsable (posuna@uacj.mx)

² USDA-ARS Jornada Experimental Range. Box 30003, NMSU, Dept. 3JER. 88003 Las Cruces, NM, USA.

Recibido: febrero de 2011. Aceptado: septiembre de 2011.

Publicado en Terra Latinoamericana 30: 39-45.

Las oleaginosas cosechadas para alimento y combustible dependen en estos cuerpos de lípidos. Las plantas también hospedan diversas poblaciones de hongos endofíticos que han escapado a la detección por el microscopio convencional. Este estudio contempla datos desde encuestas preliminares de distribución de endófitos en plantas nativas hasta examinar específicamente la asociación física entre los hongos endofíticos y los cuerpos de lípidos de las plantas. Tinciones de tejidos vegetales con azul de tripano y sudan IV a través del microscopio de interferencia diferencial de contrastes mostraron que los cuerpos de lípidos estuvieron íntimamente asociados con la hifa del hongo y con las tinciones de azul de tripano. La abundancia de los lípidos asociados con endófitos en plantas sanas sugiere que están involucrados en el metabolismo y transporte de carbono (aceite). Más investigación, particularmente a nivel molecular, es de vital importancia para entender el significado de las inclusiones de lípidos en este tipo de plantas, las cuales se conservan en pastos y matorrales. El uso de imágenes modernas y el análisis genómico o molecular ayudarían a explorar las relaciones entre las células vegetales y las células de endófitos donde se intercambian el carbón para mejorar el entendimiento del metabolismo del carbono en las plantas.

Palabras claves: zonas áridas, bioenergéticos, endófito, hongos, resistencia al estrés.

INTRODUCTION

Oils serve as a carbon and energy reserves and are precursors for membrane lipid and steroid biosynthesis in plants (Hanisch *et al.*, 2006; Demirbas, 2011). Oil or lipid bodies called oleosomes are spherical storage organelles embedded within the cytoplasm of eukaryotes (Waltermann and Steinbuechel; 2005; Martin and Parton, 2006). Oleosomes occur in cells of most plant tissues and are prominent in seeds, pollen grains, embryos and endosperm. They are also in stems, fruits and leaves, where they are frequently associated with plastids (Lersten *et al.*, 2006). The most common forms of lipids

stored in plants are triacylglycerols (TAG), triesters of fatty acids that are attached to glycerol (Moellering and Benning, 2010; Goncalves *et al.*, 2011). Oil bodies consist of a TAG matrix covered by a stabilizing layer of phospholipids and unique proteins, which are thought to originate from the endoplasmic reticulum (Goldberg *et al.*, 2009; Iwanaga *et al.*, 2008; Lersten *et al.*, 2006; Lin and Tzen, 2004). These universal components of eukaryotic cells provide a rapidly mobilized lipid source for many important biological processes (Martin and Parton, 2006; Rezanka and Sigler, 2009).

Lipid Bodies in Plant Cells

To function normally, embryos, seeds and pollen are physiologically programmed to cope with severe desiccation during maturation and storage (Liu *et al.*, 2008). Regulation of fatty acid composition may be one mechanism by which plants regulate tolerance to temperature and moisture stress (Volk *et al.*, 2007). TAGs in *Cuphea* sp. seeds is crystallized at low storage temperatures. After storage, if seeds were imbibed with water before TAGs liquefy, cells suffer irreversible membrane damage and fail to germinate. Depending upon fatty acid composition, TAGs in some species liquefy at 22 °C while other species need to be warmed to 45 °C before liquefaction and normal germination.

Oleosins, proteinaceous matrices associated with the surfaces of lipid bodies, are thought to maintain stability of lipid bodies in seeds of desiccation-tolerant plants to prevent them from coalescing during seed dehydration and germination (Murphy, 2001, Purkrtova *et al.*, 2008; Jain *et al.*, 2011). Oleosins that coat oil bodies in soybeans protect them from environmental stresses (Iwanaga *et al.*, 2008). Pollen grains characteristically contain large numbers of oil bodies that serve as energy reserves for subsequent germination (Piffanelli *et al.*, 1998; Updegraff *et al.*, 2009). Surface proteins contribute to oil body stability in lily (*Lilium longiflorum* Thumb.) pollen (Jiang *et al.*, 2007). This stability may enhance pollen viability as it is subjected to environmental extremes in transit by insects or wind. Adequate lipid storage compounds are essential for successful development during final stages of somatic embryogenesis in *Picea abies* (Grigova *et al.*, 2007).

Lersten *et al.* (2006) examined freehand cross sections of fresh leaves of 302 plant species and reported oil bodies in mesophyll cells in 71 species, but did not observe them in epidermal cells, concluding that oil

bodies are significant cellular components of many plant taxa. They found that oil bodies were common in eudicots and less frequent in monocots. They observed that in recent years oil bodies have been neglected because of a shift in plant tissue preparation. Early microscopic studies involved analysis of freehand sections that allowed vivid staining of lipids. Later methods involved killing fixatives and alcoholic dehydration that dissolved lipids and obscured their microscopic detection.

In addition to physiological roles, plant oils have wide economic importance. Two major plant oil classes are recognized: 1) volatile, low-molecular mass oils that are used commercially in perfumes and food extracts and 2) larger molecular mass, non-volatile oils that are commercially extracted from seeds of canola, castor, corn, soybean and other important crop plants (Lersten *et al.*, 2006).

Lipid Bodies Associated With Fungi

Lipid bodies structurally similar to those found in vascular plants are also common in microbes. Oleogenic fungi are those that accumulate over 25% of their dry weight as lipids when carbon is available (Murphy, 2001). Microbial oils have potential commercial value as food supplements, pharmaceuticals, and biofuels (Peng and Chen, 2007). These may be produced more economically than those extracted from agricultural crops. Peng and Chen (2007) isolated fungal endophytes with large and copious quantities of lipid bodies within their hyphae from oleagenous plants. Those with enzymatic capability to decompose wheat straw are candidates for low cost microbial oil production.

Lipid Bodies Integrated with Fungal Endophytes

Plants host diverse populations of cryptic, symbiotic fungi (Arnold *et al.*, 2000; Ganley *et al.*, 2004; Van der Heijden *et al.*, 2006; Vandenkoornhuyse *et al.*, 2002). It is well documented that fungal symbionts contribute multiple benefits that enhance tolerance to abiotic and biotic stress, but the mechanisms of inducing stress tolerance are not well understood. In our research, we have heavily utilized a dual staining approach to reveal trypan blue staining fungal chitin and Sudan IV staining lipid bodies which are associated with fungal symbionts within plant tissues (Barrow *et al.*, 2004; Barrow *et al.*, 2007; Lucero *et al.*, 2008; Barrow *et al.*, 2008; Osuna

and Barrow, 2009). We have also used molecular techniques to demonstrate the presence of complex fungal consortia that associate with plants even under aseptic conditions (Lucero *et al.*, 2008, Lucero *et al.*, 2011). Although Sudan IV stain would also reveal lipid bodies associated strictly with the plant, our collective experience suggests this is rare. Our objective in this study was to review dual stained images of plant tissues taken from studies examining endophyte distribution and other plant-fungal associations to specifically consider the association between plant lipid bodies and endophytic fungi.

MATERIALS AND METHODS

Micrographs of plant tissues retrieved from data archives of Jerry Barrow were prepared in association with several studies that occurred between 2000 and 2010. Plants used in these studies were harvested from native desert plant populations on the USDA Agricultural Research Service's Jornada Experimental Range in southwestern New Mexico, USA. Root and leaf samples were collected from actively growing grasses and shrubs. Roots described as "aseptic" were disinfected with 1.8% sodium hypochlorite solution for 10 min, and then incubated on potato dextrose agar until mycelia appeared at the root tip. Staining methods are described in Barrow and Aaltonen (2001) and Barrow (2003). Briefly, hand sectioned tissues were cleared with potassium hydroxide followed by staining with a solution containing trypan blue, which stains fungal cell walls blue, and Sudan IV, which stains lipid bodies red. Slides containing small secondary roots and leaves were mounted and examined with a Zeiss Axiophot microscope with conventional and differential interference contrast (DIC) optics at 100X and 1000X magnification. Digital images were captured and processed using Auto-Montage 3D software by Syncroscopy.

RESULTS AND DISCUSSION

Analysis of root and leaf tissues frequently revealed amorphous trypan blue stained fungal structures associated with lipid bodies on leaf surfaces (Figure 1a), root meristems (Figure 1b), lateral roots (Figure 1c) and hydrated seeds (Figure 1d) of *Bouteloua eriopoda* (1a-c), and *Sporobolus airoides* (1d), respectively. Although hyphae were sometimes visible (Figures 1a-c), fruiting bodies and other clear details useful for

identification were typically absent. Sometimes only blue stain, suggestive of fungal chitin, could be detected (Figure 1d). This amorphous fungal morphology observed among endophytes has been previously described (Barrow, 2003). Similar amorphous fungal structures, thought to be derived from yeasts, have been reported in surface sterilized seeds of *Atriplex canescens* and in micropropagated shoots of *Bouteloua eriopoda* (Osuna and Barrow, 2004) and of *Atriplex canescens*, both of which were demonstrated to contain DNA representing diverse fungal taxa (Lucero *et al.*, 2011; Lucero *et al.*, 2008).

In our studies of plants native to the Chihuahuan Desert we found extensive colonization by dark septate endophytes (DSE) (Figure 2e) (Barrow, 2003). DSE fungi are generally characterized by stained or melanized hyphae and microsclerotia (Jumpponen and Trappe, 1998), which were most prevalent in dormant plants (Figure 2a). Melanin, a natural dark pigment that makes DSE structures microscopically visible in plant tissues, was most prevalent in dormant plant tissues (mh). Lipid bodies associated with melanized hyphae in dormant plants were rare, but the abundance in physiologically active plants was remarkable (Figure 2b-d). Fungal endophytes were also detected as non-staining hyaline, clear, hyphae (Figure 1a, c, 2a, d) or with amorphous, trypan blue staining networks (Figure 1a, b, d). Lipid bodies were typically associated with all of these fungal structures.

Analysis of leaf tissues revealed lipid bodies integrated with fungal structures in photosynthetic mesophyll and bundle sheath and cells of the stomatal complex of grasses and shrubs (Barrow, 2003; Barrow and Aaltonen, 2004). In both shrub and grass species, lipid bodies were most prevalent in the root meristems, cortex and phloem. In leaves, they were most conspicuously present in photosynthetic mesophyll, bundle sheath and cells of the stomatal complex. Color images in the above cited references vividly illustrate lipid bodies associated with hyphae and other structures of endophytic fungi in tissues of native plants.

Like Lersten *et al.* (2006) we found that lipid bodies were prominent features of photosynthetic bundle sheath and mesophyll cells in many plant species. However, while Lersten *et al.* (2006) found lipid bodies to be less frequent in monocots, we observed them to be abundant if sampled during periods of high physiological activity (Figure 1, 2b-c, d). Lipid bodies were rarely observed in dormant plants (Figure 2a). Unlike Lersten *et al.* (2006)

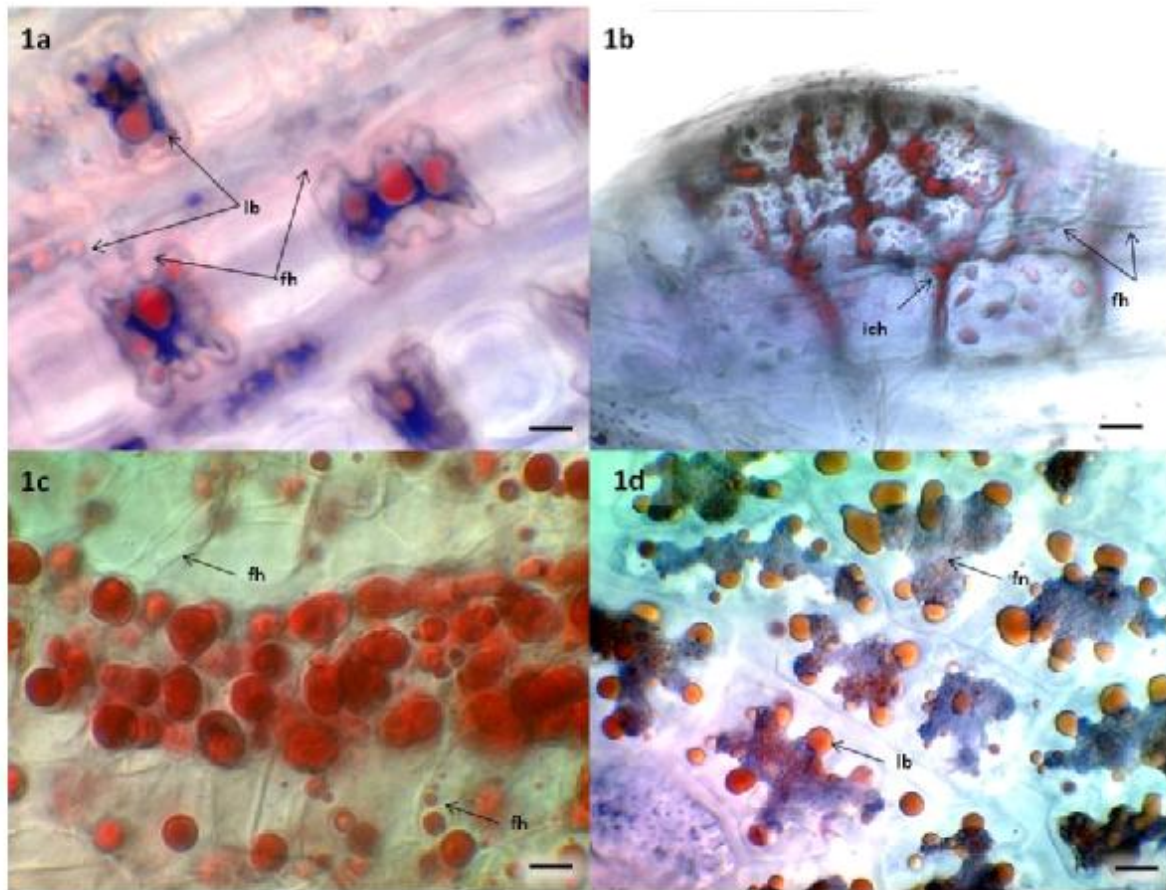


Figure 1a. Leaf mesophyll cells of *Bouteloua eriopoda*. Trypan blue- stain reveals a -hyaline branched fungal network (fh), associated with red, sudan IV stained lipid bodies (lb). Bar = 10um. **Figure 1b.** A lateral root initial of *Bouteloua eriopoda*. Sudan IV stains intercellular lipid bearing fungal protoplasts (ich) and lipid bodies (lb) within meristematic cells. Finely branched fungal hyphae (fh) and amorphous fungal structures appear blue. Bar = 10um. **Figure 1c.** Sudan IV stained lipid bodies in vascular cells of developing lateral roots of *Bouteloua eriopoda*. Hyaline fungal hyphae remain unstained and clear (fh). These hyphae surround many of the large lipid bodies near the center of the page. Lipid body- inclusions are clear in the hyaline hypha indicated with a black arrow on the lower right corner. Bar = 10um. **Figure 1d.** Endosperm cells of hydrated *Sporobolus airoides* seeds stained with trypan blue and sudan IV. A trypan blue stained fungal network (fn) with attached sudan IV stained lipid bodies (lb). Bar = 10 um.

we also observed that lipid bodies in our samples were tightly associated with endophytic fungi, which varied from finely branched networks (Figure 1a) to very dense amorphous looking structures (1c, 2b,c) (Barrow, 2003; Barrow and Aaltonen, 2004). We presume that the differences in observations are due to different research objectives (our group was primarily interested in distribution of endophytic fungi), differences in imaging (Lersten *et al.* utilized brightfield microscopy, while we utilized differential interference contrast optics to enhance clear structures), and due to our utilization of dual staining techniques. Nonetheless, we feel that highlighting this difference is important, particularly because recent interest in plant oils for biofuels, in addition to agricultural

value, is climbing. Contributions of endophytic fungi are too often overlooked in plant physiology studies. Failure to acknowledge the presence of endophytes too often leads to risky assumptions that biosynthesis, regulation, and transport of lipids and other biomolecules found within the plant is exclusively carried out by plant cells.

CONCLUSIONS

- Plant lipid bodies serve as a carbon and energy reserve for important biological processes, and are valued economically for producing food oils and biofuels. Our observations indicate that plant lipid bodies are frequently, if not always associated with endophytic fungi. This is

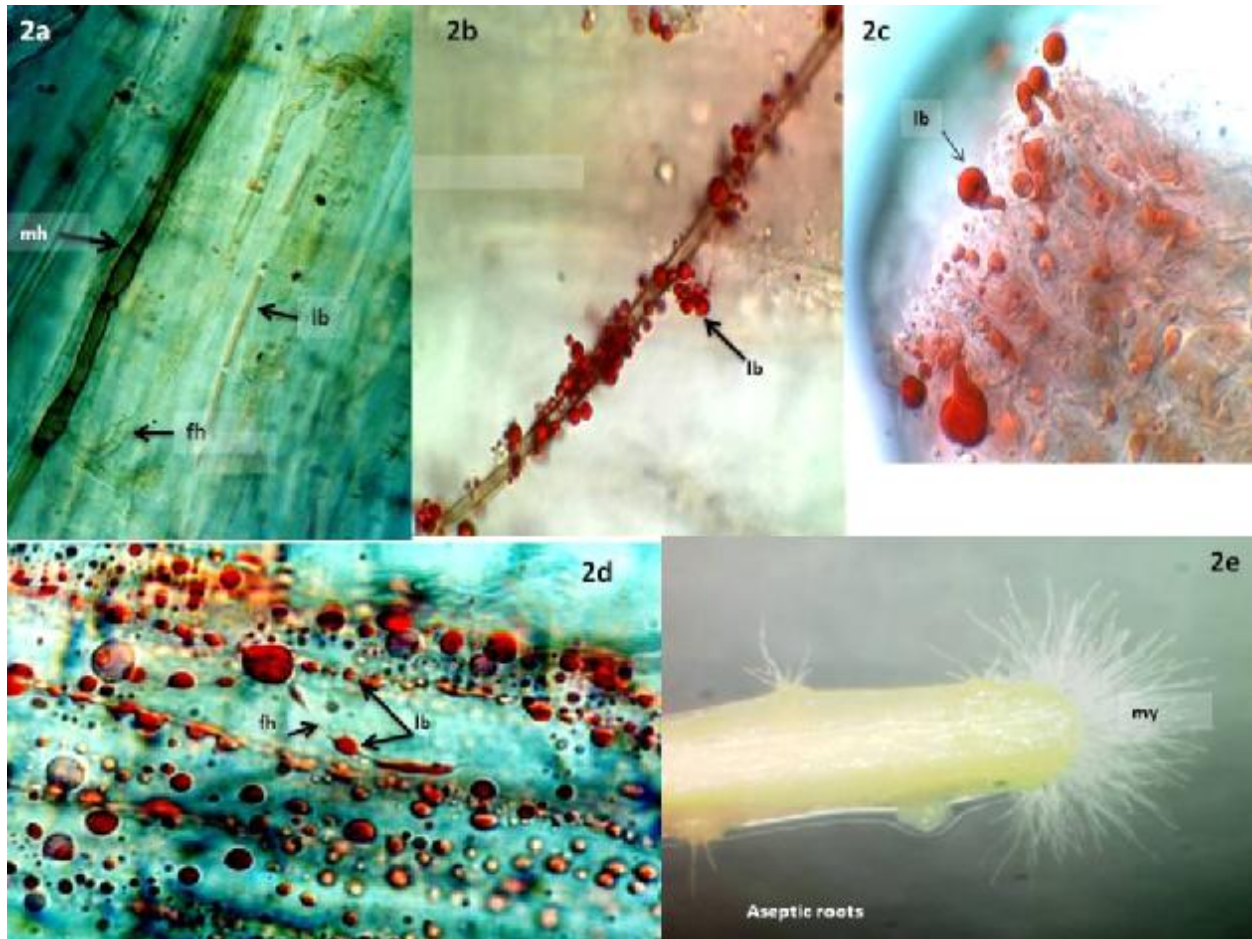


Figure 2a. Trypan blue and sudan IV stain produce limited color on dormant *B. eriopoda* roots. Brown, melanzed (mh) and fine hyaline hyphae (fh) are visible. A hyaline hypha near the center of the image contains weakly staining lipid bodies (lb) that are less developed than the lipid bodies seen during active growth stages. Figure 2b. A melanzed hypha on the surface of *B. eriopoda* is covered with lipid bodies (lb). Figure 2c. *B. eriopoda* root tissue as shown in 2e, stained to reveal lipid body droplets (lb) emerging from the zone where mycelia are visible. Figure 2d. Dual stained, physiologically active roots of *B. eriopoda* exhibit abundant intercellular lipid bodies (lb) and fine hyaline hyphae. Figure 2e. Mycelia (my) emerging from aseptate roots of *B. eriopoda* aseptate roots are visible at the tip, below the root hair zone.

novel because it highlights a largely unexplored possibility that oils in plants are produced and managed, in full or in part, by endophytic fungi.

- We propose that the physical association between lipid bodies and endophytic fungi in diverse plant species reveals a possibility that endophytic fungi may be directly involved in biosynthesis, transport, or regulation of plant oils. Considering increased global interest in accelerating production of plant oils to meet 21st century food and fuel needs, researchers exploring oil production in plants would do well to expand assessments of endophytic fungi which can play a valuable role in managing this important carbon source. More studies are necessary to explore the interfaces where plant and endophyte exchange organic carbon. Furthermore using modern imaging,

molecular and genomic analysis to assess plant interactions with associated endophyte communities could transform understanding of plant carbon metabolism.

REFERENCES

- Arnold, A. E., Z. Maynard, G. S. Gilbert, P. D. Coley, and T. A. Kursar. 2000. Are tropical fungal endophytes hyperdiverse? *Ecol. Lett.* 3:267-274.
- Barrow, J. R. 2003. Atypical morphology of dark septate fungal root endophytes of *Bouteloua* in arid southwestern USA rangelands. *Mycorrhiza* 13: 239-247.
- Barrow, J. R. and R. Aaltonen. 2004. A staining method for systemic endophytic fungi in plants. pp. 61-67. *In*: R. T. Lartey, A. J. Caesar (eds.). *Emerging concepts in plant health management*. Research Signpost, Kerala, India.

- Barrow, J. R. and R. E. Aaltonen. 2001. Evaluation of the internal colonization of *Atriplex canescens* (Pursh) Nutt. roots by dark septate fungi and the influence of host physiological activity. *Mycorrhiza* 11: 199-205.
- Barrow, J. R., M. E. Lucero, I. Reyes-Vera, and K. M. Havstad. 2008. Do symbiotic microbes have a role in plant evolution, performance and response to stress? *Commun. Integr. Biol.* 1: 69-73.
- Barrow, J. R., M. E. Lucero, I. Reyes-Vera, and K. M. Havstad. 2007. Endosymbiotic fungi structurally integrated with leaves reveals a lichenous condition of C4 grasses. *In Vitro Cell. Dev. Biol. Plant* 43: 65-70.
- Barrow, J. R.; P. Osuna-Avila, and I. Reyes-Vera. 2004. Fungal endophytes intrinsically associated with micropropagated plants regenerated from native *Bouteloua eriopoda* Torr. and *Atriplex canescens* (Pursh) Nutt. *In Vitro Cell. Dev. Biol. Plant* 40: 608-612.
- Demirbas, A. 2011. Biodiesel from oilgae, biofixation of carbon dioxide by microalgae: A solution to pollution problems. *Appl. Energ.* 88: 3541-3547.
- Facciotti, D. and V. Knauf. 1998. Triglycerides as products of photosynthesis. Genetic engineering, fatty acid composition and structure of triglycerides. pp. 225-248. *In: P. Siegenthaler and N. Murata (eds.) Lipids in photosynthesis: structure, function and genetics.* Kluwer. Dordrecht, The Netherlands.
- Ganley, R. J., S. J. Brunsfeld, and G. Newcombe. 2004. A community of unknown, endophytic fungi in western white pine. *Proc. Natl. Acad. Sci. USA* 101: 10107-10112.
- Goldberg, A. A., S. D. Bourque, P. Kyryakov, T. Boukh-Viner, C. Gregg, A. Beach, M. T. Burstein, G. Machkalyan, V. Richard, S. Rampersad, and V. I. Titorenko. 2009. A novel function of lipid droplets in regulating longevity. *Biochem. Soc. Trans.* 37: 1050-1055.
- Goncalves, K. M., F. K. Sutili, S. G. F. Leite, R. O. M. A. de Souza, and I. C. R. Leal. 2011. Palm oil hydrolysis catalyzed by lipases under ultrasound irradiation-The use of experimental design as a tool for variables evaluation. *Ultrason. Sonochem.* 19: 232-236.
- Grigova, M., M. Kubes, N. Drazna, T. Rezanka, and H. Lipavska. 2007. Storage lipid dynamics in somatic embryos of Norway spruce (*Picea abies*): histochemical and quantitative analyses. *Tree Physiol.* 27: 1533-1540.
- Hanisch, J., M. Waltermann, H. Robenek, and A. Slteinbuchel. 2006. Eukaryotic lipid body proteins in oleogenous actinomycetes and their targeting to intracellular triacylglycerol inclusions: Impact on models of lipid body biogenesis. *Appl. Environ. Microb.* 72: 6743-6750.
- Iwanaga, D., D. Gray, E. A. Decker, J. Weiss, and D. J. McClements. 2008. Stabilization of soybean oil bodies using protective pectin coatings formed by electrostatic deposition. *J. Agric. Food Chem.* 56: 2240-2245.
- Jain, M., B. P. Pathak, A. C. Harmon, B. L. Tillman, and M. Gallo. 2011. Calcium dependant protein kinase (CDPK) expression during fruit development in cultivated peanut (*Arachis hypogaea*) under Ca²⁺ sufficient and deficient growth regiments. *J. Plant Physiol.* 168: 2272-2277.
- Jiang, P. L., C. S. Wang, C. M. Hsu, G. Y. Jaub, and J. T. Tzen. 2007. Stable oil bodies sheltered by a unique oleosin in lily pollen. *Plant Cell. Physiol.* 48: 812-821.
- Jumpponen, A. and J. M. Trappe. 1998. Dark septate endophytes; a review of facultative biotrophic root-colonizing fungi. *New Phytol.* 140: 295-310.
- Lersten, N. R., A. R. Czapinski, J. D. Curtis, R. Freckmann, and H. T. Horner. 2006. Oil bodies in leaf mesophyll cells of angiosperms: overview and a selected survey. *Am. J. Bot.* 93: 1731-1739.
- Li, H.-Y., D.-W. Li, C.-M. He, Z.-P. Zhou, T. Mei, and H.-M. Xu. 2011. Diversity and heavy metal tolerance of endophytic fungi from six dominant plant species in Pb-Zn mine wasteland in China. *Fungal Ecol.* 86: 1-7.
- Lin, L. J. and J. T. Tzen. 2004. Two distinct steroleosins are present in seed oil bodies. *Plant Physiol. Biochem.* 42: 601-608.
- Liu, M. S., C. T. Chien, and T. P. Lin. 2008. Constitutive components and induced gene expression are involved in the desiccation tolerance of *Selaginella tamariscina*. *Plant Cell Physiol.* 49: 653-663.
- Lucero, M. E., A. Unc, P. Cooke, S. Dowd, and S. Sun. 2011. Endophyte Microbiome Diversity in Micropropagated *Atriplex canescens* and *Atriplex torreyi* var griffithsii. *PLoS One* 6: e17693.
- Lucero, M., J. R. Barrow, P. Osuna, and I. Reyes. 2008. A cryptic microbial community persists with in micropropagated *Bouteloua eriopoda* (Torr.) Torr. cultures. *Plant Sci.* 174: 570-575.
- Martin, S. and R. G. Parton. 2006. Lipid droplets: a unified view of a dynamic organelle. *Nat. Rev. Mol. Cell Biol.* 7: 373-378.
- Moellering, E. R. and C. Benning. 2010. RNA interference silencing of a major lipid droplet protein affects lipid droplet size in *Chlamydomonas reinhardtii*. *Eukaryot. Cell.* 9: 97-106.
- Murphy, D. J. 2001. The biogenesis and functions of lipid bodies in animals, plants and microorganisms. *Prog. Lipid Res.* 40: 325-328.
- Osuna, P. and J. R. Barrow. 2004. Regeneration of Black Grama (*Bouteloua eriopoda* Torr. Torr) Plants via somatic embryogenesis. *In Vitro Cell. Dev. Biol. Plant* 40: 299-302.
- Osuna, P. and J. R. Barrow. 2009. *In vitro* system to determine the role of *Aspergillus ustus* on *Daucus carota* roots. *Terra Latinoamericana* 27: 363-372.
- Peng, X. W. and H. Z. Chen. 2007. Microbial oil accumulation and cellulose secretion of the endophytic fungi from oleaginous plants. *Ann. Microbiol.* 57: 239-242.
- Piffanelli, P., J. H. E. Ross, and D. J. Murphy. 1998. Biogenesis and function of the lipidic structures of pollen grains. *Sex. Plant Reprod.* 11: 65-80.
- Purkrtova, Z., P. Jolivet, M. Miquel, and T. Chardot. 2008. Structure and function of seed lipid-body-associated proteins. *C. R. Biol.* 331: 746-754.
- Rezanka, T. and K. Sigler. 2009. Odd-numbered very-long-chain fatty acids from microbial, animal and kingdoms. *Prog. Lipid Res.* 48: 206-238.
- Updegraff, E. P., F. Zhao, and D. Preuss. 2009. The extracellular lipase EXL4 is required for efficient hydration of Arabidopsis pollen. *Sex. Plant Reprod.* 22: 197-204.
- Van der Heijden, M. G., R. Bakker, J. Verwaal, T. R. Scheublin, M. Ruppen, R. Van Logtestijn, and C. Staehelin. 2006. Symbiotic bacteria as a determinant of plant community structure and plant productivity in dune grassland. *FEMES Microbiol. Ecol.* 56: 178-187.

- Vandenkoornhuise, P., S. L. Baldauf, C. Leyval, J. Straczek, and J. P.W. Young. 2002. Extensive fungal diversity in plant roots. *Science* 295: 2051.
- Volk, G. M., J. Crane, A. M. Caspersen, D. A. Kovach, C. A. Gardner, and C. T. Walters. 2007. Hydration of *Cuphea* seeds containing crystallized triacylglycerols. *Funct. Plant Biol.* 34: 360-367.
- Waltermann, M. and A. Steinbüchel. 2005. Neutral lipid bodies in prokaryotes: Recent insights into structure formation, and relationship to eukaryotic lipid depots. *J. Bacteriol.* 187: 3607-3619.